



# ATLANTIC TESTING LABORATORIES

**WBE certified company**

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May 28, 2024

Boquet Valley Central School District  
7530 Court Street  
Elizabethtown, New York 12932

Attn: Josh Meyer

Re: Air Sampling and Analysis Services  
Boquet Valley CSD Indoor Air Quality  
Lake View Campus, Westport, New York  
Mountain View Campus, Elizabethtown, New York  
Essex County, New York  
ATL Report No. PL6065IH-01-05-24

Ladies/Gentlemen:

Enclosed is a copy of our report for the Air Sampling and Analysis Services performed at the referenced sites. This project was completed in accordance with the scope of work outlined in our contract (ATL No. PL5998-199-04-24), dated April 4, 2024, and authorized by Josh Meyer on April 6, 2024.

Please contact our office should you have any questions, or if we may be of further assistance.

Sincerely,  
*ATLANTIC TESTING LABORATORIES, Limited*

Robert B. Read  
Project Manager

RBR/CJD/jdf

Enclosures

**AIR SAMPLING AND ANALYSIS SERVICES**

**BOQUET VALLEY CSD INDOOR AIR QUALITY  
LAKE VIEW CAMPUS, WESTPORT, NEW YORK  
MOUNTAIN VIEW CAMPUS, ELIZABETHTOWN, NEW YORK**



*WBE certified company*

**PREPARED FOR:**

**Boquet Valley Central School District  
7530 Court Street  
Elizabethtown, New York 12932**

**PREPARED BY:**

**Atlantic Testing Laboratories, Limited  
130 Arizona Avenue, Suite 1540  
Plattsburgh, New York 12903**

**ATL REPORT NO. PL6065IH-01-05-24**

**MAY 28, 2024**

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## **1.0 INTRODUCTION**

Atlantic Testing Laboratories, Limited (ATL) was retained to provide air sampling and analysis services for the Lake View and Mountain View Campuses of the Boquet Valley Central School District, spanning the towns of Westport and Elizabethtown, in Essex County, New York. The purpose of the air sampling and analysis services was to evaluate potential adverse air quality impacts with respect to common general air quality parameters within the school buildings, in response to recent observations made by building occupants. Services were provided in accordance with the scope of work in our contract (ATL No. PL5998-199-04-24, dated April 4, 2024).

## **2.0 FIELD OBSERVATIONS**

The subject buildings currently function as school buildings. Each of the school buildings generally consists of 2 stories and a basement. Interior building materials within the subject building include, but may not be limited to, floor tile, carpet, gypsum board, plaster, ceiling tile, wood, and concrete. The Mountain View Campus was constructed in 1958, and the Lake View Campus was constructed in 1933.

## **3.0 INDOOR AIR QUALITY MONITORING**

### **3.1 Monitoring Locations and Methodology**

Indoor air quality monitoring was conducted using a TSI Q-Trak Model 7575 indoor air quality monitor. This instrument provides real-time measurements for carbon dioxide, carbon monoxide, relative humidity, and temperature. Indoor air quality monitoring for the measurable presence of airborne particles, ranging from 0.3 micrometers (um) to 10 um, was performed using a Fluke 985 particle counter. Indoor air quality monitoring for the measurable presence of volatile organic compounds (VOC) was conducted using a RAE 3000+ portable photoionization detector (PID). The RAE 3000+ PID is equipped with a 10.6eV lamp and has a range of 1 part per billion (ppb) to 20,000 parts per million (ppm). The PID was calibrated in accordance with the manufacturer's specifications prior to use.

Areas that were sampled using the indoor air quality monitor, particle counter, and PID were selected to provide representative data throughout selected rooms of the subject building. Sample measurements were collected within the approximate breathing zone (4 to 6 feet above the floor). At the time of the monitoring event, the sampled areas were occupied.

### **3.2 Summary of Monitoring Data**

The results of the indoor air quality monitoring activities for carbon monoxide, carbon dioxide, relative humidity, temperature, and VOC are provided in Table B-1, of Attachment B. The results of the indoor air quality monitoring for airborne particles are provided in Table B-2, of Attachment B. Findings of the indoor air quality monitoring are further discussed in Section 5.

## **4.0 MOLD SAMPLING AND ANALYSIS**

### **4.1 General Information about Mold**

Molds are simple, microscopic organisms that can be found almost anywhere. Molds can grow on virtually any organic substance, provided there is moisture and oxygen present. There are

molds that have the capability to grow on wood, paper, carpet, food, insulation, and numerous other products and building materials. When excessive moisture accumulates in buildings or on building materials, mold growth will often occur, particularly if the moisture problem remains undiscovered or unresolved. It is impossible to eliminate all molds and mold spores in the indoor environment; however, controlling the amount of moisture within the building can reduce the potential for mold propagation.

The reproduction of molds involves the creation of microscopic spores that usually cannot be seen without magnification. Due to the size of mold spores, these can easily become airborne. Airborne mold spores can be either viable or non-viable. Although the non-viable mold spores do not have the ability to reproduce, these spores can be as equally detrimental to an individual's health as the viable mold spores. Viable airborne mold spores are of major concern, because of the ability to reproduce, in addition to the potential health hazards that may be created. When viable airborne mold spores come into contact with a damp surface, the spores typically begin to reproduce and form a conglomeration of mold spores. The conglomeration will digest the material upon which it is growing, and will continue to reproduce at a rate that varies depending on the type of mold, quantity of moisture present, the amount of food source available, and other environmental factors, including the temperature and humidity levels of the surroundings. Molds gradually damage the objects grown on, consequently posing a threat to the structural integrity of a building over time.

There are numerous types of molds that exist in the environment. Although certain types of mold have been shown to severely affect people, it is important to note that all molds have the potential to cause health effects. Molds can produce allergens that may trigger allergic reactions or asthma attacks, and certain types of molds are known to produce potent toxins and/or irritants. People that may be affected more severely include infants and children, elderly individuals, pregnant women, individuals with respiratory conditions or allergies and asthma, and persons with weakened immune systems. Potential health concerns are an important reason to prevent mold growth and to remediate any existing indoor mold growth.

## 4.2 Spore Classifications

Results for samples that are laboratory analyzed for mold spores are typically reported as different fungal spore classifications. The laboratory utilized for this project, Galson Laboratories, has differentiated between 13 classifications for air samples. A brief description of the spore classifications is provided below. The descriptions identify typical reported characteristics for each classification, and are provided for informational purposes. This information is not intended to represent an exact scientific evaluation.

1. *Alternaria*: The *Alternaria* group contains approximately 40 to 50 species, and is usually more prevalent during the summer and early fall months. This type of mold grows rapidly and is a known source of allergens in the atmosphere, generally causing symptoms associated with respiratory problems. *Alternaria* is one of the most common fungi worldwide, and typically grows in soil, dead organic debris, food, and textiles.
2. *Ascospores*: *Ascospores* are found everywhere in nature and are predominantly forcibly discharged during periods of high humidity or rain. *Ascospores* are extremely variable in size and shape, and a vast majority is reported to not cause diseases in humans. The laboratory designation for *Ascospores* includes all ascospores with the exception of *Chaetomium*.

3. *Aspergillus/Penicillium-like*: The *Aspergillus* and *Penicillium* species are commonly found in indoor environments. The *Aspergillus* species exist worldwide, and typically grow in soil, decayed vegetation, and other kinds of organic matter. Only a few of these molds have been reported to cause disease in humans; however, this type has the ability to produce mycotoxins. The *Penicillium* species are common contaminants that are found on various substrates. Many species of *Penicillium* are known potential mycotoxin producers; however, human pathogenic species are rare. This type of mold can pose a danger indoors because of the capability to grow and reproduce in just a few days. The laboratory analysis groups *Aspergillus* and *Penicillium* together as the spores are indistinguishable on a spore trap.
4. *Basidiospores*: *Basidiospores* develop from mushrooms and wood decay fungi, and are abundant in the environment. *Basidiospores* are frequently detected at high levels in the outside air, and may grow indoors under suitable conditions. Although *Basidiospores* are not known to produce mycotoxins in the traditional sense, large numbers of airborne *Basidiospores* can be allergenic, and some forms may cause rare opportunistic infections.
5. *Bipolaris/Drechslera*: The *Bipolaris* and *Drechslera* species are ubiquitous, cosmopolitan species that grow on a variety of substrates. Colonies are observed as shades of dark gray to brown. These spores are the most commonly reported cause of allergic fungal sinusitis. Production of toxins by these species is currently unknown. The laboratory designation for *Bipolaris/Drechslera* includes *Helminthosporium* and *Exserohilium*.
6. *Chaetomium*: *Chaetomium* is a common fungus that is distributed worldwide and typically found in soil, decaying organic matter, seeds, wood, and other cellulose-containing materials. Mold spores for *Chaetomium* are relatively large and settle from the air more quickly than other mold types. This mold type frequently emits a musty odor and is considered impossible to remediate without removal of the impacted materials. *Chaetomium* is not a common pathogen in humans and is usually not considered a major concern unless disturbed.
7. *Cladosporium*: The *Cladosporium* group contains over 500 species. These species are widely distributed in air and rotten organic material. This type of mold can pose a danger indoors, because of the capability to grow and reproduce in just a few days. It is frequently found in elevated levels in water-damaged environments, and is only occasionally associated with disease in humans.
8. *Curvularia*: The *Curvularia* group consists of approximately 30 species. These are most commonly found in tropical and subtropical regions; however, a few species do exist in the temperate zones. *Curvularia* species are a common cause of allergic reactions, and may cause infections in humans.
9. *Mycelial Fragments*: *Mycelial Fragments* are the dead and decaying fragments from fungi, molds, and yeast. Although *Mycelial Fragments* do not have the ability to reproduce, these provide a food source for other mold types and can continue to adversely affect the health of humans if inhaled or ingested.
10. *Pollen*: *Pollen* is a fine to coarse powder necessary for plant reproduction. *Pollen* grains have a hard coat for protection during movement. *Pollen* is a very common allergen and typically causes seasonal "hay fever" allergies in susceptible persons.
11. *Rusts/Smuts*: *Rusts/Smuts* are parasitic plant pathogens that require a living host for growth, and therefore, do not grow indoors unless the host plants are present. Spores from

*Rusts/Smuts* may cause allergic reactions, but are not reported to be infectious to humans. The laboratory designation for Rusts/Smuts includes *Myxomycetes* and *Periconia*.

12. **Stachybotrys**: *Stachybotrys* is a greenish-black mold that is one of the most widely known mold types, due to its existence in many high profile mold-infested buildings and the potential for this type of mold to produce extremely potent toxins. Although many reports of toxicity effects on humans from exposure to this fungus are anecdotal, reported health effects have included cold and flu symptoms, memory loss, muscle aches, sore throats, headaches, fatigue, dermatitis, intermittent local hair loss, cancer, and generalized malaise. The ability of this fungus to produce toxins depends on the material it is growing on and environmental conditions, such as temperature, pH, and humidity. Since it is impossible to control all of these factors to prevent the production of toxins, it is usually assumed that *Stachybotrys*, if present, is toxic to the surrounding environment. *Stachybotrys* requires very wet or high humid conditions for days or weeks in order to grow; however, once this mold begins to grow, it has the capability to continue to propagate without the existence of a water source, consequently making it difficult to detect and remediate all impacted areas. Furthermore, due to the size of the spores and composition of a *Stachybotrys* mold colony, *Stachybotrys* mold spores are not readily released into the air, and detection of even a few of these spores in an air sample usually indicates that *Stachybotrys* has started to colonize somewhere in very high numbers. The laboratory designation for *Stachybotrys* includes *Memnoniella*.
13. **Other/Unidentified**: *Other* spores are those observed on the spore trap that can be identified, but are rarely observed and/or are typically observed in small quantities. Unidentified includes broken and dehydrated spores, spores partially obscured by debris, and spores that can't be categorized solely with microscopy.

### 4.3 Air Samples

#### 4.3.1 Sampling Locations and Methodology

Air sampling was conducted using Zefon Air-O-Cell™ cassettes, provided by Galson Laboratories. The Zefon Air-O-Cell™ is an impaction-based air sampler designed to pull air across a tacky sampling medium, trapping any airborne particulates. A high-flow vacuum pump, set at approximately 15 liters per minute, is utilized in conjunction with the Zefon Air-O-Cell™ to extract air from the immediate surroundings and pull this air across the tacky sampling medium. The procedure for collecting air samples using the Zefon Air-O-Cell™ includes attaching tubing and adapters that connect the high-flow vacuum pump with an Air-O-Cell™ cassette. The high-flow vacuum pump and Air-O-Cell™ cassette are then positioned in the area that is selected for sampling. After setup is complete and the tubing and connections are checked to ensure proper airflow, the vacuum pump is activated for a specified duration. Air samples collected for this project were set at 5-minute durations. This duration is typically used for an average room with minimum visible dirt, as recommended by the supplier of the sampling media.

During the sampling event, a total of 19 air samples were collected at representative locations, including samples taken at building entrances to serve as backgrounds. Table 1 provides a summary of the air sample identifications and locations.

**Table 1**  
**Summary of Air Samples**

Sample ID	Sample Date	Sample Location	Sample Type
<b>Mountainview Campus</b>			
PL6065MA-01	April 25, 2024	Old Library	Interior area of concern
PL6065MA-02	April 25, 2024	Crawlspace	Interior area of concern
PL6065MA-03	April 25, 2024	Gym	Interior area of concern
PL6065MA-04	April 25, 2024	Room No. 112	Interior area of concern
PL6065MA-05	April 25, 2024	Room No. 120	Interior area of concern
PL6065MA-06	April 25, 2024	Room No. 217	Interior area of concern
PL6065MA-07	April 25, 2024	Room No. 210	Interior area of concern
PL6065MA-08	April 25, 2024	Principal's Office	Interior area of concern
PL6065MA-09	April 25, 2024	Front Entrance	Entrance Background
PL6065MA-10	April 25, 2024	Gym Entrance	Entrance Background
<b>Lakeview Campus</b>			
PL6065MA-11	April 26, 2024	Room No. 121	Interior area of concern
PL6065MA-12	April 26, 2024	Cafeteria	Interior area of concern
PL6065MA-13	April 26, 2024	Room No. 001	Interior area of concern
PL6065MA-14	April 26, 2024	Room No. 101	Interior area of concern
PL6065MA-15	April 26, 2024	Room No. 125	Interior area of concern
PL6065MA-16	April 26, 2024	Room No. 217	Interior area of concern
PL6065MA-17	April 26, 2024	Room No. 214	Interior area of concern
PL6065MA-18	April 26, 2024	Room No. 203	Interior area of concern
PL6065MA-19	April 26, 2024	Exterior	Entrance Background

After collecting and properly securing the 19 air samples, the Air-O-Cell™ cassettes were returned to Galson Laboratories for analysis.

#### **4.3.2 Summary of Laboratory Data**

A copy of the laboratory report, including sample custody documentation, is contained in Appendix A. A summary of the mold analytical results for the collected air samples is provided in Table C-1 and C-2 of Appendix C.

Classification of the fungal spores into 13 different types was completed for each air sample, as indicated in the laboratory report and Table C-1 and C-2 in Appendix C. Additional information provided in the laboratory reports includes the percent composition of each of the 13 classifications relative to the whole sample, and the estimated crowding factor for each of the air samples.

The estimated crowding factor provides a relative quantification of the density of particles contained within the Air-O-Cell™ cassettes that may interfere with the spore counts. The crowding factor is rated on a scale of 0 to 5, with 0 corresponding to no particles detected and 5 corresponding to an overcrowding of particles of such a magnitude as to render analysis impossible. With the exception of sample MA02, each of the collected air samples from the sampling event had a crowding factor of 2. This crowding factor corresponds to samples exhibiting particles that are close together and overlapping, and the spore counts are potentially biased low. Air sample MA02 had a crowding factor of 4. This crowding factor corresponds to samples exhibiting particles that are very crowded, and the spore counts are likely to be biased low.



Data interpretation for air samples is generally based on the comparison of indoor and outdoor spore counts. There are currently no guidelines or regulations to indicate "safe" or "normal" spore levels; however, typical indoor counts are 30 to 80 percent of outdoor spore counts, with the same general distribution of spore types present. Variation is also an inherent part of biological air sampling. The presence or absence of a few genera in small numbers typically is not considered abnormal.

The total spore count concentration for an indoor sample should naturally be lower than that of an outdoor sample. If this condition is not satisfied, there is a strong possibility that the mold spores contained within the building are being generated by a source other than the natural interaction with the outside environment. Specifically, in buildings without excessive presence of mold, the qualitative diversity of airborne fungi indoors and outdoors should be similar. Conversely, the dominating presence of one or two kinds of fungi indoors and the absence of the same kind outdoors may be indicative of a moisture problem and/or degraded indoor air quality.

For periods of cold climatic conditions, comparison between indoor and outdoor mold spore classifications is complicated by the significantly reduced presence of airborne mold spores in the outdoor environment. For these colder climatic conditions, indoor mold spore count concentrations reported at levels higher than the outdoor may not necessarily represent an immediate concern. Such conditions need to be evaluated relative to the actual reported concentrations for the indoor air samples (i.e., significantly high concentrations detected) and the types of mold spores detected (i.e., common types found indoors or less common and possibly more toxic types). Furthermore, additional sampling events during warmer periods may be beneficial for confirmation of findings.

Tables C-1 and C-2 in Appendix C identifies the variations between indoor and outdoor samples that were collected for this project. A further explanation of these variations is provided in Section 5 of this report.

## **5.0 DISCUSSION OF FINDINGS**

The following list of findings is presented as a generalized summary of the results and observations provided during performance of the air quality monitoring for the Mountain View Campus and Lake View Campus of the Boquet Valley CSD, Essex County, New York.

1. Average relative humidity measurements in the subject buildings ranged from 14% to 38%. Although there are no regulated guidance values for indoor relative humidity, industry wide recommendations suggest an average indoor relative humidity of 40% to 60%. Although, at the time of sampling, the relative humidity was measured to be dryer than the recommended range, ambient outdoor relative humidity was measured from 10% to 22% and may indicate that the measured indoor values may not be representative of typical conditions. While lower indoor relative humidity is beneficial relative to mold, a medium range indoor relative humidity is often more beneficial relative to airborne-transmitted infections, bacteria, and viruses. A higher relative humidity does not directly produce adverse indoor air quality, but could provide favorable conditions for different airborne particulates/contaminants. It is not anticipated that the indoor relative humidity levels would need to be further assessed or addressed at this time.
2. Carbon monoxide is a toxic, colorless, odorless, and combustible gas that is a product of incomplete combustion. It is generated by many sources, such as gasoline-powered internal combustion engines, arc welding (used as an inert gas), and fires. The

Occupational Safety and Health Administration (OSHA) have promulgated a permissible exposure limit of 50 parts per million (ppm) carbon monoxide for a time-weighted-average (TWA) 8-hour work shift. Several of the symptoms of carbon monoxide exposure include headaches, tachypnea, nausea, dizziness, and cyanosis. A detectable level (1.4 ppm) of carbon monoxide was detected at the front entrance of the Mountain View campus, but at a level well below the OSHA permissible limits. Carbon monoxide was not detected at any of the other locations tested, as identified in Section 3.2 of this report, and is therefore not currently considered a concern.

3. Carbon dioxide is a colorless, odorless gas that is a normal constituent of air (approximately 250 to 500 ppm) and is produced in the respiration process of living beings. The burning of fossil fuels is an additional source. The symptoms of carbon dioxide exposure are similar to carbon monoxide.

Measurement of carbon dioxide levels within an occupied building is a method for ascertaining the adequacy of the ventilation system. The National Institute for Occupational Safety and Health (NIOSH) indicates that levels greater than 800 ppm suggest the ventilation system may be inadequate. The American Society of Heating, Refrigerating, and Air-Conditioning Engineers (ASHRAE) recommend adjusting the building's ventilation system when carbon dioxide levels exceed 1,000 ppm. When sufficient outdoor air is supplied to keep carbon dioxide levels below 1,000 ppm, the ventilation is generally considered to be adequate. The presence of increased levels of carbon dioxide in the indoor air of buildings is generally attributed to occupancy. Increased levels can also be indicative of an inefficient ventilation system. Elevated carbon dioxide concentrations in a building reflect insufficient exchange of "fresh" outdoor air for "spent" interior air, allowing the accumulation of carbon dioxide and possibly other contaminants.

Average carbon dioxide levels recorded in the selected sample locations, as identified in Table A-1 of Attachment A, ranged from 400 ppm to 1,700 ppm. Of the 16 measurements at the Mountain View Campus, 7 were above 800 ppm and 3 were above 1,000 ppm. For the Lake View Campus, 7 of the 37 measurements were above 800 ppm and 5 were above 1,000 ppm. With the guidelines provided by NIOSH and ASHRAE, the current ventilation system may need further assessment to verify adequate air exchange throughout the building.

4. New building materials and paints, adhesives, stains, etc. can increase the level of volatile organic compounds (VOC). Other common sources of VOC for a building include cleaning products and occupant induced fragrances (i.e., perfume, cologne, air fresheners).

A PID was utilized to collect real-time VOC concentrations throughout the subject building. No detectable levels of VOC were encountered during the sampling event. Indoor VOC are not currently considered to be a concern in reference to the air quality of the subject buildings.

5. Particle counts, as measured by field instrumentation and summarized in Table B-2 of Attachment A, show various locations where the indoor measurements were significantly higher than the outdoor locations. As indicated in Item 3, the ventilation system may need further assessment to verify adequate air exchange and filtration throughout the buildings. If there are areas with visible dust, cleaning or more frequent cleaning of these areas may also reduce airborne particulate.

6. Air sampling for subsequent mold analysis included the collection of 16 samples within the buildings during the sampling events. Each air sampling event also included samples from outside the building to serve as background samples and provide a standard for comparison.

Based on a review of the laboratory analysis results for individual spore classifications, exceedances were identified for interior air samples collected.

- a. Air Sample PL6065MA01, collected from the Mountain View Old Library exhibited a concentration of the *Other/Unidentified* spore type at a level exceeding the corresponding concentration for the background sample.
- b. Air Sample PL6065MA02, collected from the Mountain View Crawlspace, exhibited concentrations of the *Mycelial Fragments*, *Aspergillus/Penicillium-like*, *Chaetomium*, *Cladosporium*, *Rust/Smuts*, and *Other/Unidentified* spore types at levels exceeding the corresponding concentration for the background sample.
- c. Air Sample PL6065MA03, collected from the Mountain View Gymnasium, exhibited concentrations of the *Mycelial Fragments*, *Aspergillus/Penicillium-like*, and *Cladosporium* spore types at levels exceeding the corresponding concentration for the background sample.
- d. Air Sample PL6065MA04, collected from the Mountain View Room No. 112, exhibited concentrations of the *Mycelial Fragments*, *Total Fungal Spores*, *Cladosporium*, *Curvularia*, *Rust/Smuts*, and *Other/Unidentified* spore types at levels exceeding the corresponding concentration for the background sample.
- e. Air Sample PL6065MA05, collected from the Mountain View Room No. 120, exhibited concentrations of the *Mycelial Fragments*, *Total Fungal Spores*, *Aspergillus/Penicillium-like*, *Basidiospores*, *Cladosporium*, and *Other/Unidentified* spore types at levels exceeding the corresponding concentration for the background sample.
- f. Air Sample PL6065MA06, collected from the Mountain View Room No. 217, exhibited concentrations of the *Mycelial Fragments*, *Total Fungal Spores*, *Aspergillus/Penicillium-like*, *Basidiospores*, *Cladosporium*, *Curvularia*, and *Other/Unidentified* spore types at levels exceeding the corresponding concentration for the background sample.
- g. Air Sample PL6065MA07, collected from the Mountain View Room No. 210, exhibited concentrations of the *Mycelial Fragments*, *Total Fungal Spores*, and *Cladosporium* spore types at levels exceeding the corresponding concentration for the background sample.
- h. Air Sample PL6065MA08, collected from the Mountain View Principal Office, exhibited concentrations of the *Alternaria* and *Cladosporium* types at levels exceeding the corresponding concentration for the background sample.
- i. Air Sample PL6065MA11, collected from the Lake View Room No. 121, exhibited concentrations of the *Mycelial Fragments*, *Pollen*, *Total Fungal Spores*, *Basidiospores*, *Cladosporium*, and *Other/Unidentified* spore types at levels exceeding the corresponding concentration for the background sample.
- j. Air Sample PL6065MA12, collected from the Lake View Room Cafeteria, exhibited concentrations of the *Total Fungal Spores*, *Aspergillus/Penicillium-like*, *Basidiospores*,

*Cladosporium*, and *Other/Unidentified* spore types at levels exceeding the corresponding concentration for the background sample.

- k. Air Sample PL6065MA13, collected from the Lake View Room No. 001, exhibited concentrations of the *Mycelial Fragments*, *Pollen*, *Total Fungal Spores*, and *Basidiospores* spore types at levels exceeding the corresponding concentration for the background sample.
- l. Air Sample PL6065MA14, collected from the Lake View Room No. 101, exhibited concentrations of the *Mycelial Fragments*, *Pollen*, *Total Fungal Spores*, *Aspergillus/Penicillium-like*, *Basidiospores*, *Cladosporium*, and *Other/Unidentified* spore types at levels exceeding the corresponding concentration for the background sample.
- m. Air Sample PL6065MA15, collected from the Lake View Room No. 125, exhibited concentrations of the *Pollen* and *Aspergillus/Penicillium-like* spore types at levels exceeding the corresponding concentration for the background sample.
- n. Air Sample PL6065MA16, collected from the Lake View Room No. 217, exhibited concentrations of the *Mycelial Fragments* and *Cladosporium* spore types at levels exceeding the corresponding concentration for the background sample.
- o. Air Sample PL6065MA17, collected from the Lake View Room No. 214, exhibited concentrations of the *Mycelial Fragments*, *Pollen*, *Total Fungal Spores*, *Basidiospores*, and *Cladosporium* spore types at levels exceeding the corresponding concentration for the background sample.
- p. Air Sample PL6065MA18, collected from the Lake View Room No. 203, exhibited concentrations of the *Mycelial Fragments*, *Pollen*, *Total Fungal Spores*, *Aspergillus/Penicillium-like*, *Basidiospores*, *Cladosporium*, *Rust/Smuts*, and *Other/Unidentified* spore types at levels exceeding the corresponding concentration for the background sample.

Information available from the sampling and analysis events is not indicative of a clear and obvious adverse impact to indoor air quality relative to mold spores; however, the noted exceedances for some of the indoor air samples further support efforts for assessment of the ventilation system and more frequent cleaning of areas that may be dusty or exhibit the higher airborne particulate counts.

- 7. The sampling services that were provided for this project included the collection of data for relatively short durations of time, rather than monitoring the air quality continuously. Consequently, the field data and analytical results are valid only for that specific period of time for the selected instrument and location of measurement. It is possible for indoor air quality parameters to change dramatically over time, and these may even change on a daily basis, depending on the usage of the building, the efficiency of the HVAC system, the introduction of adverse conditions, and various other factors. Future monitoring, sampling, and/or analysis of areas within the subject building may need to be considered if there is a substantial alteration in the building usage, or if additional factors contributing to air quality issues become prevalent.

**ATTACHMENT A**

**LABORATORY REPORT AND SAMPLE CUSTODY DOCUMENTATION**



**GALSON**

**Robert Read  
Atlantic Testing Laboratories  
130 Arizona Ave  
Plattsburgh, NY 12903**

**May 01, 2024**

**Account# 12293**

**Login# L624585**

**Dear Robert Read:**

**Enclosed are the analytical results for the samples received by our laboratory on April 29, 2024. All samples on the chain of custody were received in good condition unless otherwise noted. Any additional observations will be noted on the chain of custody.**

**Please contact client services at (888) 432-5227 if you would like any additional information regarding this report. Thank you for using SGS Galson.**

**Sincerely,**

**SGS Galson**

A handwritten signature in black ink that reads 'Lisa Swab'. The signature is written in a cursive, flowing style.

**Lisa Swab  
Laboratory Director**

**Enclosure(s)**



**Terms and Conditions & General Disclaimers**

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**Analytical Disclaimers**

- Unless otherwise noted within the report, all quality control results associated with the samples were within established control limits or did not impact reported results.
- Note: The findings recorded within this report were drawn from analysis of the sample(s) provided to the laboratory by the Client (or a third party acting at the Client’s direction). The laboratory does not have control over the sampling process, including but not limited to the use of field equipment and collection media, as well as the sampling duration, collection volume or any other collection parameter used by the Client. The findings herein constitute no warranty of the sample's representativeness of any sampled environment, and strictly relate to the samples as they were presented to the laboratory. For recommended sampling collection parameters, please refer to the Sampling and Analysis Guide at [www.sgsgalson.com](http://www.sgsgalson.com).
- Unrounded results are carried through the calculations that yield the final result and the final result is rounded to the number of significant figures appropriate to the accuracy of the analytical method. Please note that results appearing in the columns preceding the final result column may have been rounded and therefore, if carried through the calculations, may not yield an identical final result to the one reported.
- The stated LOQs for each analyte represent the demonstrated LOQ concentrations prior to correction for desorption efficiency (if applicable).
- Unless otherwise noted within the report, results have not been blank corrected for any field blank or method blank data.

**Accreditations** SGS Galson holds a variety of accreditations and recognitions. Our quality management system conforms with the requirements of ISO/IEC 17025. Where applicable, samples may also be analyzed in accordance with the requirements of ELAP, NELAC, or LELAP under one of the state accrediting bodies listed below. Current Scopes of Accreditation can be viewed at <http://www.sgsgalson.com> in the accreditations section of the "About" page. To determine if the analyte tested falls under our scope of accreditation, please visit our website or call Client Services at (888) 432-5227.

National/International	Accreditation/Recognition	Lab ID#	Program/Sector
AIHA-LAP, LLC - IHLAP, ELLAP, EMLAP	ISO/IEC 17025 and USEPA NLLAP	Lab ID 100324	Industrial Hygiene, Environmental Lead, Environmental Microbiology

State	Accreditation/Recognition	Lab ID#	Program/Sector
New York (NYSDOH)	ELAP and NELAC (TNI)	Lab ID: 11626	Air Analysis, Solid and Hazardous Waste
Louisiana (LDEQ)	LELAP	Lab ID: 04083	Air Analysis, Solid Chemical Materials

**Legend**

< - Less than	mg - Milligrams	MDL - Method Detection Limit	ppb - Parts per Billion
> - Greater than	ug - Micrograms	NA - Not Applicable	ppm - Parts per Million
l - Liters	m3 - Cubic Meters	NS - Not Specified	ppbv - ppb Volume
LOQ - Limit of Quantitation	kg - Kilograms	ND - Not Detected	ppmv - ppm Volume
ft2 - Square Feet	cm2 - Square Centimeters	in2 - Square Inches	ng - Nanograms



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Client : Atlantic Testing Laboratories  
 Site : MOUNTAIN VIEW LAKE VIEW  
 Project No. : BOQUET VALLEY CSD  
 Date Sampled : 25-APR-24 - 26-APR-24  
 Date Received : 29-APR-24  
 Incubation Temp : NA

Account No.: 12293  
 Login No. : L624585  
 Date Analyzed : 01-MAY-24  
 Report ID : 1421391

Client ID : PL6065MA01    Lab ID : L624585-1    Air Volume : 0.075 m3  
 Analysis : Standard Mold Screen    Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	3	3	40	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	13	13	170	NA
-----				
<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium-like</i>	2	2	27	15.4
Basidiospores	5	5	67	38.5
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	2	2	27	15.4
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	4	4	53	30.8

**COMMENTS:** Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore    Submitted by: TAC/SLS    Supervisor: BDB    Date : 01-MAY-24  
 Analytical Method : In-house: IB-AIROCELL; Mic    Approved by : BDB    Sampler : Spore Trap

CFU -Colony Forming Units    g -Grams





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 Date Received : 29-APR-24  
 Incubation Temp : NA

Account No.: 12293  
 Login No. : L624585  
 Date Analyzed : 01-MAY-24  
 Report ID : 1421391

Client ID : PL6065MA02      Lab ID : L624585-2      Air Volume : 0.075 m3  
 Analysis : Standard Mold Screen      Crowding Factor : 4

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	46	46	610	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	75	75	1000	NA
-----				
<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	2	2	27	2.7
<i>Aspergillus/Penicillium</i> -like	17	17	230	22.7
Basidiospores	9	9	120	12
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	20	20	270	26.7
<i>Cladosporium</i>	7	7	93	9.3
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	6	6	80	8
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	14	14	190	18.7

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore      Submitted by: TAC/SLS      Supervisor: BDB      Date : 01-MAY-24  
 Analytical Method : In-house: IB-AIROCELL; Mic      Approved by : BDB      Sampler : Spore Trap

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 Date Received : 29-APR-24  
 Incubation Temp : NA

Account No.: 12293  
 Login No. : L624585  
 Date Analyzed : 01-MAY-24  
 Report ID : 1421391

Client ID : PL6065MA03      Lab ID : L624585-3      Air Volume : 0.075 m3  
 Analysis : Standard Mold Screen      Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	7	7	93	NA
Pollen	2	2	27	NA
Total Fungal Spores	19	19	250	NA
-----				
<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	1	1	13	5.3
<i>Aspergillus/Penicillium</i> -like	4	4	53	21.1
Basidiospores	8	8	110	42.1
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	4	4	53	21.1
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	2	2	27	10.5

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore      Submitted by: TAC/SLS      Supervisor: BDB      Date : 01-MAY-24  
 Analytical Method : In-house: IB-AIROCELL; Mic      Approved by : BDB      Sampler : Spore Trap

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 Date Sampled : 25-APR-24 - 26-APR-24  
 Date Received : 29-APR-24  
 Incubation Temp : NA

Account No.: 12293  
 Login No. : L624585  
 Date Analyzed : 01-MAY-24  
 Report ID : 1421391

Client ID : PL6065MA04      Lab ID : L624585-4      Air Volume : 0.075 m3  
 Analysis : Standard Mold Screen      Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	9	9	120	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	40	40	530	NA
-----				
<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	1	1	13	2.5
<i>Aspergillus/Penicillium</i> -like	2	2	27	5
Basidiospores	13	13	170	32.5
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	12	12	160	30
<i>Curvularia</i>	1	1	13	2.5
Rusts/Smuts	6	6	80	15
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	5	5	67	12.5

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore      Submitted by: TAC/SLS      Supervisor: BDB      Date : 01-MAY-24  
 Analytical Method : In-house: IB-AIROCELL; Mic      Approved by : BDB      Sampler : Spore Trap

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 Date Sampled : 25-APR-24 - 26-APR-24  
 Date Received : 29-APR-24  
 Incubation Temp : NA

Account No.: 12293  
 Login No. : L624585  
 Date Analyzed : 01-MAY-24  
 Report ID : 1421391

Client ID : PL6065MA05      Lab ID : L624585-5      Air Volume : 0.075 m3  
 Analysis : Standard Mold Screen      Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	4	4	53	NA
Pollen	1	1	13	NA
Total Fungal Spores	42	42	560	NA
-----				
<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium</i> -like	7	7	93	16.7
Basidiospores	19	19	250	45.2
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	5	5	67	11.9
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	2	2	27	4.8
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	9	9	120	21.4

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore      Submitted by: TAC/SLS      Supervisor: BDB      Date : 01-MAY-24  
 Analytical Method : In-house: IB-AIROCELL; Mic      Approved by : BDB      Sampler : Spore Trap

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 Date Sampled : 25-APR-24 - 26-APR-24  
 Date Received : 29-APR-24  
 Incubation Temp : NA

Account No.: 12293  
 Login No. : L624585  
 Date Analyzed : 01-MAY-24  
 Report ID : 1421391

Client ID : PL6065MA06      Lab ID : L624585-6      Air Volume : 0.075 m3  
 Analysis : Standard Mold Screen      Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	9	9	120	NA
Pollen	1	1	13	NA
Total Fungal Spores	56	56	750	NA
-----				
<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	1	1	13	1.8
<i>Aspergillus/Penicillium</i> -like	12	12	160	21.4
Basidiospores	18	18	240	32.1
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	9	9	120	16.1
<i>Curvularia</i>	1	1	13	1.8
Rusts/Smuts	4	4	53	7.1
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	11	11	150	19.6

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore      Submitted by: TAC/SLS      Supervisor: BDB      Date : 01-MAY-24  
 Analytical Method : In-house: IB-AIROCELL; Mic      Approved by : BDB      Sampler : Spore Trap

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 Date Sampled : 25-APR-24 - 26-APR-24  
 Date Received : 29-APR-24  
 Incubation Temp : NA

Account No.: 12293  
 Login No. : L624585  
 Date Analyzed : 01-MAY-24  
 Report ID : 1421391

Client ID : PL6065MA07      Lab ID : L624585-7      Air Volume : 0.075 m3  
 Analysis : Standard Mold Screen      Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	11	11	150	NA
Pollen	1	1	13	NA
Total Fungal Spores	38	38	510	NA
-----				
<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	2	2	27	5.3
<i>Aspergillus/Penicillium</i> -like	1	1	13	2.6
Basidiospores	13	13	170	34.2
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	19	19	250	50
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	2	2	27	5.3
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	1	1	13	2.6

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore      Submitted by: TAC/SLS      Supervisor: BDB      Date : 01-MAY-24  
 Analytical Method : In-house: IB-AIROCELL; Mic      Approved by : BDB      Sampler : Spore Trap

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 Date Received : 29-APR-24  
 Incubation Temp : NA

Account No.: 12293  
 Login No. : L624585  
 Date Analyzed : 01-MAY-24  
 Report ID : 1421391

Client ID : PL6065MA08      Lab ID : L624585-8      Air Volume : 0.075 m3  
 Analysis : Standard Mold Screen      Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	5	5	67	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	20	20	270	NA
-----				
<i>Alternaria</i>	1	1	13	5
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium</i> -like	2	2	27	10
Basidiospores	7	7	93	35
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	6	6	80	30
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	1	1	13	5
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	3	3	40	15

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore      Submitted by: TAC/SLS      Supervisor: BDB      Date : 01-MAY-24  
 Analytical Method : In-house: IB-AIROCELL; Mic      Approved by : BDB      Sampler : Spore Trap

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 Date Sampled : 25-APR-24 - 26-APR-24  
 Date Received : 29-APR-24  
 Incubation Temp : NA

Account No.: 12293  
 Login No. : L624585  
 Date Analyzed : 01-MAY-24  
 Report ID : 1421391

Client ID : PL6065MA09      Lab ID : L624585-9      Air Volume : 0.075 m3  
 Analysis : Standard Mold Screen      Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	5	5	67	NA
Pollen	1	1	13	NA
Total Fungal Spores	27	27	360	NA
-----				
<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	2	2	27	7.4
<i>Aspergillus/Penicillium</i> -like	2	2	27	7.4
Basidiospores	14	14	190	51.9
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	2	2	27	7.4
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	4	4	53	14.8
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	3	3	40	11.1

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore      Submitted by: TAC/SLS      Supervisor: BDB      Date : 01-MAY-24  
 Analytical Method : In-house: IB-AIROCELL; Mic      Approved by : BDB      Sampler : Spore Trap

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 Date Received : 29-APR-24  
 Incubation Temp : NA

Account No.: 12293  
 Login No. : L624585  
 Date Analyzed : 01-MAY-24  
 Report ID : 1421391

Client ID : PL6065MA10      Lab ID : L624585-10      Air Volume : 0.075 m3  
 Analysis : Standard Mold Screen      Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	2	2	27	NA
Pollen	4	4	53	NA
Total Fungal Spores	21	21	280	NA
-----				
<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	1	1	13	4.8
<i>Aspergillus/Penicillium-like</i>	1	1	13	4.8
Basidiospores	15	15	200	71.4
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	1	1	13	4.8
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	2	2	27	9.5
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	1	1	13	4.8

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore      Submitted by: TAC/SLS      Supervisor: BDB      Date : 01-MAY-24  
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 Date Received : 29-APR-24  
 Incubation Temp : NA

Account No.: 12293  
 Login No. : L624585  
 Date Analyzed : 01-MAY-24  
 Report ID : 1421391

Client ID : PL6065MA11      Lab ID : L624585-11      Air Volume : 0.075 m3  
 Analysis : Standard Mold Screen      Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	8	8	110	NA
Pollen	1	1	13	NA
Total Fungal Spores	27	27	360	NA
-----				
<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	1	1	13	3.7
<i>Aspergillus/Penicillium</i> -like	1	1	13	3.7
Basidiospores	18	18	240	66.7
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	3	3	40	11.1
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	4	4	53	14.8

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore      Submitted by: TAC/SLS      Supervisor: BDB      Date : 01-MAY-24  
 Analytical Method : In-house: IB-AIROCELL; Mic      Approved by : BDB      Sampler : Spore Trap

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 Date Received : 29-APR-24  
 Incubation Temp : NA

Account No.: 12293  
 Login No. : L624585  
 Date Analyzed : 01-MAY-24  
 Report ID : 1421391

Client ID : PL6065MA12      Lab ID : L624585-12      Air Volume : 0.075 m3  
 Analysis : Standard Mold Screen      Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	2	2	27	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	28	28	370	NA
-----				
<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium</i> -like	5	5	67	17.9
Basidiospores	16	16	210	57.1
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	2	2	27	7.1
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	1	1	13	3.6
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	4	4	53	14.3

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore      Submitted by: TAC/SLS      Supervisor: BDB      Date : 01-MAY-24  
 Analytical Method : In-house: IB-AIROCELL; Mic      Approved by : BDB      Sampler : Spore Trap

CFU -Colony Forming Units      g -Grams



# GALSON

## LABORATORY ANALYSIS REPORT

6601 Kirkville Road  
 East Syracuse, NY 13057  
 (315) 432-5227  
 FAX: (315) 437-0571  
 www.sgsgalson.com

Client : Atlantic Testing Laboratories  
 Site : MOUNTAIN VIEW LAKE VIEW  
 Project No. : BOQUET VALLEY CSD  
 Date Sampled : 25-APR-24 - 26-APR-24  
 Date Received : 29-APR-24  
 Incubation Temp : NA

Account No.: 12293  
 Login No. : L624585  
 Date Analyzed : 01-MAY-24  
 Report ID : 1421391

Client ID : PL6065MA13      Lab ID : L624585-13      Air Volume : 0.075 m3  
 Analysis : Standard Mold Screen      Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	7	7	93	NA
Pollen	1	1	13	NA
Total Fungal Spores	23	23	310	NA
-----				
<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	2	2	27	8.7
<i>Aspergillus/Penicillium</i> -like	1	1	13	4.3
Basidiospores	18	18	240	78.3
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	1	1	13	4.3
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	1	1	13	4.3

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore      Submitted by: TAC/SLS      Supervisor: BDB      Date : 01-MAY-24  
 Analytical Method : In-house: IB-AIROCELL; Mic      Approved by : BDB      Sampler : Spore Trap

CFU -Colony Forming Units      g -Grams



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Client : Atlantic Testing Laboratories  
 Site : MOUNTAIN VIEW LAKE VIEW  
 Project No. : BOQUET VALLEY CSD  
 Date Sampled : 25-APR-24 - 26-APR-24  
 Date Received : 29-APR-24  
 Incubation Temp : NA

Account No.: 12293  
 Login No. : L624585  
 Date Analyzed : 01-MAY-24  
 Report ID : 1421391

Client ID : PL6065MA14      Lab ID : L624585-14      Air Volume : 0.075 m3  
 Analysis : Standard Mold Screen      Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	7	7	93	NA
Pollen	1	1	13	NA
Total Fungal Spores	35	35	470	NA
-----				
<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	3	3	40	8.6
<i>Aspergillus/Penicillium</i> -like	6	6	80	17.1
Basidiospores	14	14	190	40
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	8	8	110	22.9
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	1	1	13	2.9
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	3	3	40	8.6

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore      Submitted by: TAC/SLS      Supervisor: BDB      Date : 01-MAY-24  
 Analytical Method : In-house: IB-AIROCELL; Mic      Approved by : BDB      Sampler : Spore Trap

CFU -Colony Forming Units      g -Grams



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## LABORATORY ANALYSIS REPORT

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Client : Atlantic Testing Laboratories  
 Site : MOUNTAIN VIEW LAKE VIEW  
 Project No. : BOQUET VALLEY CSD  
 Date Sampled : 25-APR-24 - 26-APR-24  
 Date Received : 29-APR-24  
 Incubation Temp : NA

Account No.: 12293  
 Login No. : L624585  
 Date Analyzed : 01-MAY-24  
 Report ID : 1421391

Client ID : PL6065MA15      Lab ID : L624585-15      Air Volume : 0.075 m3  
 Analysis : Standard Mold Screen      Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	2	2	27	NA
Pollen	1	1	13	NA
Total Fungal Spores	13	13	170	NA
-----				
<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	1	1	13	7.7
<i>Aspergillus/Penicillium</i> -like	4	4	53	30.8
Basidiospores	7	7	93	53.8
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	1	1	13	7.7
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	<1	<1	<13	NA

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore      Submitted by: TAC/SLS      Supervisor: BDB      Date : 01-MAY-24  
 Analytical Method : In-house: IB-AIROCELL; Mic      Approved by : BDB      Sampler : Spore Trap

CFU -Colony Forming Units      g -Grams



# GALSON

## LABORATORY ANALYSIS REPORT

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 www.sgsgalson.com

Client : Atlantic Testing Laboratories  
 Site : MOUNTAIN VIEW LAKE VIEW  
 Project No. : BOQUET VALLEY CSD  
 Date Sampled : 25-APR-24 - 26-APR-24  
 Date Received : 29-APR-24  
 Incubation Temp : NA

Account No.: 12293  
 Login No. : L624585  
 Date Analyzed : 01-MAY-24  
 Report ID : 1421391

Client ID : PL6065MA16      Lab ID : L624585-16      Air Volume : 0.075 m3  
 Analysis : Standard Mold Screen      Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	3	3	40	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	19	19	250	NA
-----				
<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	1	1	13	5.3
<i>Aspergillus/Penicillium</i> -like	3	3	40	15.8
Basidiospores	11	11	150	57.9
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	3	3	40	15.8
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	<1	<1	<13	NA
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	1	1	13	5.3

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore      Submitted by: TAC/SLS      Supervisor: BDB      Date : 01-MAY-24  
 Analytical Method : In-house: IB-AIROCELL; Mic      Approved by : BDB      Sampler : Spore Trap

CFU -Colony Forming Units      g -Grams



# GALSON

## LABORATORY ANALYSIS REPORT

6601 Kirkville Road  
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Client : Atlantic Testing Laboratories  
 Site : MOUNTAIN VIEW LAKE VIEW  
 Project No. : BOQUET VALLEY CSD  
 Date Sampled : 25-APR-24 - 26-APR-24  
 Date Received : 29-APR-24  
 Incubation Temp : NA

Account No.: 12293  
 Login No. : L624585  
 Date Analyzed : 01-MAY-24  
 Report ID : 1421391

Client ID : PL6065MA17      Lab ID : L624585-17      Air Volume : 0.075 m3  
 Analysis : Standard Mold Screen      Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	13	13	170	NA
Pollen	3	3	40	NA
Total Fungal Spores	29	29	390	NA
-----				
<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	<1	<1	<13	NA
<i>Aspergillus/Penicillium</i> -like	1	1	13	3.4
Basidiospores	16	16	210	55.2
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	10	10	130	34.5
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	1	1	13	3.4
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	1	1	13	3.4

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore      Submitted by: TAC/SLS      Supervisor: BDB      Date : 01-MAY-24  
 Analytical Method : In-house: IB-AIROCELL; Mic      Approved by : BDB      Sampler : Spore Trap

CFU -Colony Forming Units      g -Grams





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Client : Atlantic Testing Laboratories  
 Site : MOUNTAIN VIEW LAKE VIEW  
 Project No. : BOQUET VALLEY CSD  
 Date Sampled : 25-APR-24 - 26-APR-24  
 Date Received : 29-APR-24  
 Incubation Temp : NA

Account No.: 12293  
 Login No. : L624585  
 Date Analyzed : 01-MAY-24  
 Report ID : 1421391

Client ID : PL6065MA18      Lab ID : L624585-18      Air Volume : 0.075 m3  
 Analysis : Standard Mold Screen      Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	14	14	190	NA
Pollen	2	2	27	NA
Total Fungal Spores	46	46	610	NA
-----				
<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	1	1	13	2.2
<i>Aspergillus/Penicillium</i> -like	6	6	80	13
Basidiospores	17	17	230	37
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	6	6	80	13
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	4	4	53	8.7
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	12	12	160	26.1

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore      Submitted by: TAC/SLS      Supervisor: BDB      Date : 01-MAY-24  
 Analytical Method : In-house: IB-AIROCELL; Mic      Approved by : BDB      Sampler : Spore Trap

CFU -Colony Forming Units      g -Grams



# GALSON

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 www.sgsgalson.com

Client : Atlantic Testing Laboratories  
 Site : MOUNTAIN VIEW LAKE VIEW  
 Project No. : BOQUET VALLEY CSD  
 Date Sampled : 25-APR-24 - 26-APR-24  
 Date Received : 29-APR-24  
 Incubation Temp : NA

Account No.: 12293  
 Login No. : L624585  
 Date Analyzed : 01-MAY-24  
 Report ID : 1421391

Client ID : PL6065MA19      Lab ID : L624585-19      Air Volume : 0.075 m3  
 Analysis : Standard Mold Screen      Crowding Factor : 2

Parameter	Raw Count	Total Count	Conc Count/m3	Percent %
Mycelial Fragments	2	2	27	NA
Pollen	<1	<1	<13	NA
Total Fungal Spores	21	21	280	NA
-----				
<i>Alternaria</i>	<1	<1	<13	NA
Ascospores	3	3	40	14.3
<i>Aspergillus/Penicillium</i> -like	3	3	40	14.3
Basidiospores	12	12	160	57.1
<i>Bipolaris/Drechslera</i>	<1	<1	<13	NA
<i>Chaetomium</i>	<1	<1	<13	NA
<i>Cladosporium</i>	1	1	13	4.8
<i>Curvularia</i>	<1	<1	<13	NA
Rusts/Smuts	1	1	13	4.8
<i>Stachybotrys</i>	<1	<1	<13	NA
Other/Unidentified	1	1	13	4.8

COMMENTS: Please see attached lab footnote report for any applicable footnotes.

Level of Quantitation: 1 Spore      Submitted by: TAC/SLS      Supervisor: BDB      Date : 01-MAY-24  
 Analytical Method : In-house: IB-AIROCELL; Mic      Approved by : BDB      Sampler : Spore Trap

CFU -Colony Forming Units      g -Grams



# GALSON

LABORATORY FOOTNOTE REPORT

6601 Kirkville Road  
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FAX: (315) 437-0571  
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Client Name : Atlantic Testing Laboratories  
Site : MOUNTAIN VIEW LAKE VIEW  
Project No. : BOQUET VALLEY CSD

Date Sampled : 25-APR-24 - 26-APR-24 Account No.: 12293  
Date Received: 29-APR-24 Login No. : L624585  
Date Analyzed: 01-MAY-24

---

L624585 (Report ID: 1421391):  
SOPs: ib-airocell(29)

L624585-2 (Report ID: 1421391):  
Due to excessive debris on sample, some fungi may not have been detected.



6601 Kirkville Road  
 East Syracuse, NY 13057-0369  
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<http://www.sgsgalson.com>

## Analytical Notes for Microbiology Air-O-Cell™ Cassettes and other Spore Traps

Air-O-Cell™ cassettes and other spore traps may capture non-microbial particles that may interfere with spore counts. SGS Galson provides an estimation of the density of these particles, referred to as a Crowding Factor. The Crowding Factor ranges from 0 to 5 and is explained below. High levels of particulate matter on the impaction medium may bias the analysis by obscuring or covering spores. In addition, particle capture efficiency may decrease with high levels of particulate matter.

Crowding Factor	Explanation
0	No particles detected. This is typical of blank samples. Because most air samples typically contain some particles, absence of particulate matter could indicate improper sampling if the sample was not meant to be a blank.
1	Particles are far apart and in low numbers. Particulate matter covers approximately <5% of the impaction area. Spore counts not affected or minimally affected by the particle load.
2	Particles are close together and/or overlapping, and some spores may be obscured. Particulate matter covers approximately 5% to 25% of the impaction area. Spore counts may be biased low.
3	Particles are moderately crowded. It is likely that some spores are obscured. Particulate matter covers approximately 25% to 75% of the impaction area. Spore counts are likely biased low.
4	Particles are crowded, frequently obscuring spores. Particulate matter covers approximately 75% to 90% of the impaction area. Spore counts are likely biased low. The degree of bias increases with the percent of the trace that is occluded.
5	Particles are overcrowded making analysis impossible; no spore counts provided. If certain spores are readily detectable, they are reported as "Detected". If heavy quantities of spores are observed along the edges of the trace, this is footnoted in the report.



Counts for any genus that exceed 300 spores are estimated to two significant figures.

The list of fungal spores reported is:

***Alternaria* includes spores previously reported as *Ulocladium*.**

**Ascospores** – includes all ascospores with the exception of *Chaetomium*.

***Aspergillus/Penicillium*-like** – These two genera are grouped together as the spores are indistinguishable on a spore trap.

**Basidiospores** – This includes all basidiospores, even ones that can be identified to genus level, such as *Ganoderma*.

***Bipolaris/Drechslera*** – *Helminthosporium* and *Exserohilum* are included in this grouping.

***Chaetomium*** – Due to its unique shape and due to the fact that it may be associated with indoor mold problems, this ascospore is reported separate from other ascospores.

***Cladosporium***

***Curvularia***

**Rusts/Smuts** – *Myxomycetes* and *Periconia* are included in this grouping.

***Stachybotrys*** – This includes *Memnoniella*.

***Ulocladium* has been reclassified and is now reported as *Alternaria***

**Other/Unidentified** – “Other” includes spores that can be identified but are rarely observed and/or are typically seen in small quantities. They include: *Acremonium*, *Botrytis*, *Cercospora*, *Epicoccum*, *Fusarium*, *Nigrospora*, *Oidium*, *Paecilomyces*, *Pestalotia*, *Pestalotiopsis*, *Pithomyces*, *Polythrincium*, *Scopulariopsis*, *Spegazzinia*, *Stemphylium*, *Taeniocella*, *Tetraploa*, *Torula*, and *Trichoderma*, and *Zygophiala*. “Unidentified” includes broken and dehydrated spores, spores that are partially obscured by debris, and spores that can't be categorized using microscopy alone.

In addition, other analytes that will be shown on reports include mycelial fragments (hyphae) and pollen.

Reports for expanded analysis include the above list with the addition of skin cells and fibers.

Generally, 100% of the sample deposit is analyzed. However, some analytes with high counts may be estimated based on the analysis of a portion of the slide and the results extrapolated. In these cases, the reported values will differ between the “Raw Count” and “Total Count” columns. For example, if an analyst observed 304 basidiospores after analyzing 25% of the sample, the estimated value is 1216. The final report would show 304 in the “Raw Count” column and 1200 in the “Total Count” column (the “Total Column” is rounded to two significant figures).



## **Direct Microscopic Examination (Screens)**

- The analytes that we report are the same as those listed for spore traps with the exceptions of pollen, skin cells, and fibers.
- Due to the inherent nature of screen samples, a spore count is not performed.
- Upon special request counts may be performed on swab, liquid, or bulk screens. Counts are never performed on tape lifts due to the nature of the samples to not have uniform distribution of spores.
- The amount of a particular spore detected is reported as a “Level of contamination”. The level of contamination is a subjective measurement and corresponds to the general quantity of spores present in a sample. It also describes the amount of spores relative to one another.
  - Light: approximately 1 to 5 spores or mycelial fragments per microscope field of view at 600x.
  - Moderate: 6 to 15 spores or mycelial fragments per microscope field of view at 600x.
  - Heavy: Greater than 15 spores or mycelial fragments per microscope field of view at 600x.

## **Viable Fungi Analysis**

- Standard growing conditions for viable fungi are  $25^{\circ}\text{C} \pm 1^{\circ}\text{C}$  for 7 days.
- Standard growing conditions for viable thermophilic fungi are  $37^{\circ}\text{C} \pm 1^{\circ}\text{C}$  for 7 days.
- Results are reported in colony forming units (CFUs). A CFU can originate from one or many spores.
- SGS Galson uses and provides Potato Dextrose agar for all cultureable fungal methods. We have found Potato Dextrose agar to be suitable for the culture of the widest range of organisms. Other agars submitted or requested by clients are grown under the above standard conditions unless otherwise requested by the client.
- Some fungi may not produce identifiable structures in culture or under standard growing conditions. These fungi will be considered sterile hyphae and reported as such.
- Lack of growth under standard conditions does not preclude the presence of fungi or its viability in a sample.
- Samples taken with impactor samplers are not corrected for a positive hole correction factor.
- Identification of fungal organisms is based on visual microscopic examination at up to seven days of growth under standard conditions. Due to the large numbers of different species that may comprise them, certain genera may appear similar due to variations in stages of their life cycles, growth requirements, and/or environmental stress. A very limited amount of identification overlap may occur due to morphological similarities.
- Final interpretation of results is up to the person(s) responsible for conducting the sampling.



## Quality Control/Quality Assurance

- A daily quality control spore trap slide is read each day that an analyst performs analysis on client spore trap samples. These slides consist of old client samples that have been analyzed a minimum of twenty times before they are used as a part of the quality control program. Control limits are set at the mean plus or minus three standard deviations for each analyte and for the total spore count. Warning limits are set at the mean plus or minus two standard deviations for each analyte and for the total spore count.
- A minimum of five percent of the samples are analyzed as duplicates and five percent of the samples are analyzed as replicates (or at least one replicate or duplicate per day). The relative percent difference (RPD) is calculated between the original sample result and its duplicate or replicate. The RPD value must fall within statistically based limits. In addition, there must be agreement between three of the top five categories.
- Daily quality control includes a blind spore trap challenge and a blind fungal culture identification challenge. Each analyst must correctly identify a spore or other airborne particulate from an old spore trap slide and identify a slide prepared from a fungal culture, respectively.
- Monthly quality control includes quantifying and identifying a viable culture to genus level.
- Prior to analyzing samples, each microscope's Kohler illumination is checked. The microscope fields of view are calibrated annually.
- The lactophenol dye, slides, cover slips and spore traps are checked on a daily basis to assure that there is no contamination. Upon initial receipt, one spore trap from each lot that SGS Galson receives is checked for possible contamination.
- Media used for viable analysis is tested upon receipt for both sterility and growth promotion.
- A second analyst reexamines samples that have no observable spores.
- All reports undergo a secondary quality assurance review prior to release.

121043179065927723

Date: 04/29/24

Shipper: UPS

Initials: MMM



Prep: UNKNOWN

U024580

113

# CHAIN OF CUSTODY

<input checked="" type="checkbox"/> Standard 0% <input type="checkbox"/> 4 Business Days 35% <input type="checkbox"/> 3 Business Days 50% <input type="checkbox"/> 2 Business Days 75% <input type="checkbox"/> Next Day by 6pm 100% <input type="checkbox"/> Next Day by Noon 150% <input type="checkbox"/> Same Day 200%	Client Acct No.: 12293	Report To: Robert Read	Invoice To: Accounts Payable
	Original Prep No.: PSY736957	Company Name: Atlantic Testing Laboratories	Company Name: Atlantic Testing Laboratories
	Online COC No.: 295382	Address 1: 130 Arizona Ave	Address 1: 6431 US Highway 11
		Address 2:	Address 2: P.O. Box 29
		City, State Zip: Plattsburgh, NY 12903	Company Name: Canton, NY 13617
		Phone No.: 518-563-5878	Phone No.: 315-386-4578
		Cell No.:	Email Address: ap@atlantictesting.com
	Email reports to: rread@atlantictesting.com, labsAT@atlantictesting.com	Comments:	
	Email EDD to: rread@atlantictesting.com, labsAT@atlantictesting.com	P.O. No.: PL6065	
	Comments:	Payment info: <input type="checkbox"/> I will call SGS to provide credit card info <input type="checkbox"/> Card on File (enter the last five digits on the line below)	

Comments: \_\_\_\_\_ State Sampled: NY  MSHA

Site Name: Mountain View & Lake View Project: Boquet Valley CSD Sampled By: R. Read List description of industry or Processes/Interfaces present in sampling area: Schools

Sample ID (Maximum of 20 Characters)	Date Sampled	Collection Medium	Sample Volume Sample Time Sample Area	Liters Minutes in <sup>2</sup> , cm <sup>2</sup> , ft <sup>2</sup>	Analysis Requested	Method Reference	Internal Notes
PL6065MA01	4/25/2024	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	MV Old Library
PL6065MA02	4/25/2024	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	MV Crawford
PL6065MA03	4/25/2024	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	MV Gym

If the method(s) indicated on the COC are not our routine/preferred method(s), we will substitute our routine/preferred methods. If this is not acceptable, check here to have us contact you.

Chain of Custody	Print Name / Signature	Date	Time	Print Name / Signature	Date	Time
Relinquished By:	<u>Robert Read</u>	<u>4/26/2024</u>	<u>1645</u>	Received By: <u>Megan M. McGrath</u>	<u>4/29/24</u>	<u>12:02</u>
Relinquished By:				Received By:		

Samples received after 3pm will be considered as next day's business.

Online COC No.: 295382  
Prep No.: PSY736957  
Account No.: 12293  
Finalized: 04/23/2024 12:38:19

All services are rendered in accordance with the applicable SGS General Conditions of Service accessible via: <http://www.sgs.com/en/Terms-and-Conditions.aspx>





# CHAIN OF CUSTODY

Comments:

Sample ID (Maximum of 20 Characters)	Date Sampled	Collection Medium	Sample Volume Sample Time Sample Area	Liters Minutes in <sup>2</sup> , cm <sup>2</sup> , ft <sup>2</sup>	Analysis Requested	Method Reference	Internal Notes
PL6065MA04	4/25/2024	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	MV 112
PL6065MA05	4/25/2024	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	MV 120
PL6065MA06	4/25/2024	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	MV 217
PL6065MA07	4/25/2024	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	MV 216
PL6065MA08	4/25/2024	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	MV Principal
PL6065MA09	4/25/2024	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	MV Front Entrance
PL6065MA10	4/25/2024	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	MV Gym Entrance
PL6065MA11	4/25/2024	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	LV 121
PL6065MA12	4/25/2024	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	LV 067eria
PL6065MA13	4/25/2024	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	LV 001

If the method(s) indicated on the COC are not our routine/preferred method(s), we will substitute our routine/preferred methods. If this is not acceptable, check here to have us contact you.

Chain of Custody	Print Name / Signature	Date	Time	Print Name / Signature	Date	Time
Relinquished By:	Robert Reed	4/26/2024	1645	Received By: Megan M. McGrath	4/29/24	12:02
Relinquished By:	GA 6U			Received By:		

Samples received after 3pm will be considered as next day's business.

Online COC No. :295382

Prep No. :PSY736957

Account No. :12293

Finalized :04/23/2024 12:38:19

All services are rendered in accordance with the applicable SGS General Conditions of Service accessible via: <http://www.sgs.com/en/Terms-and-Conditions.aspx>



# CHAIN OF CUSTODY

Comments:							
Sample ID (Maximum of 20 Characters)	Date Sampled	Collection Medium	Sample Volume Sample Time Sample Area	Liters Minutes in <sup>2</sup> , cm <sup>2</sup> , ft <sup>2</sup>	Analysis Requested	Method Reference	Internal Notes
PL6065MA14	4/26/2024	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	LV 101
PL6065MA15	4/26/2024	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	LV 125
PL6065MA16	4/26/2024	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	LV 217
PL6065MA17	4/26/2024	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	LV 214
PL6065MA18	4/26/2024	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	LV 203
PL6065MA19	4/26/2024	Air-O-Cell	75	L	Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	LV Entrance
		<del>Air-O-Cell</del>			Standard Mold Screen	In-house: IB-AIROCELL; Microscopy	(RBR)

If the method(s) indicated on the COC are not our routine/preferred method(s), we will substitute our routine/preferred methods. If this is not acceptable, check here to have us contact you.

Chain of Custody	Print Name / Signature	Date	Time	Print Name / Signature	Date	Time
Relinquished By:	Robert Reed	4/26/2024	1645	Received By: Megan M. McGrath	4/29/24	12:02
Relinquished By:				Received By:		

Samples received after 3pm will be considered as next day's business.

Online COC No. :295382  
 Prep No. :PSY736957  
 Account No. :12293  
 Finalized :04/23/2024 12:38:19

All services are rendered in accordance with the applicable SGS General Conditions of Service accessible via: <http://www.sgs.com/en/Terms-and-Conditions.aspx>

## **APPENDIX B**

### **SUMMARY OF COLLECTED INDOOR AIR QUALITY DATA**

**Table B-1**  
**Summary of Air Monitoring Data (CO, CO<sub>2</sub>, Temperature, Relative Humidity, VOC)**

Location/Room	CO (ppm)	CO <sub>2</sub> (ppm)	Temperature (°F)	Relative Humidity (%)	VOC (ppb)
<b>Mountain View Campus</b>					
Old Library	0.0	760	66.1	38.1	0
Kitchen/Cafeteria	0.0	553	70.1	29.1	0
Crawlspace	0.0	539	71.5	25.9	0
Gym/Locker/Weight	0.0	557	71.0	23.4	0
112	0.0	665	71.2	22.2	0
120	0.0	942	69.8	20.5	0
217	0.0	1,645	71.2	31.4	0
218	0.0	995	71.2	27.3	0
210	0.0	775	69.5	27.5	0
Second Floor Hall	0.0	1,019	70.0	28.3	0
226	0.0	1,378	71.4	29.4	0
116	0.0	955	70.5	25.5	0
Principal	0.0	987	70.0	22.6	0
507	0.0	629	72.1	23.3	0
Front Entrance	1.4	424	50.1	16.4	0
Gym Entrance	0.0	440	46.9	21.5	0
<b>Lake View Campus</b>					
Library 121	0.0	681	65.2	26.9	0
Boiler	0.0	592	73.0	19.5	0
Shop 003	0.0	495	74.0	19.1	0
Music 017	0.0	857	74.3	21.2	0
025	0.0	750	74.9	16.9	0
Cafeteria	0.0	551	71.7	19.1	0
004	0.0	766	73.0	20.6	0
New Gym	0.0	485	68.2	20.8	0
001 OT/PT	0.0	564	67.0	22.3	0
007	0.0	1,322	68.3	27.6	0
009	0.0	1,202	69.6	26.7	0
100	0.0	675	70.7	24.3	0
102	0.0	860	71.8	25.4	0
101	0.0	789	74.1	24.6	0
103	0.0	696	74.3	22.2	0
Gym 105	0.0	620	73.6	20.2	0
113	0.0	670	74.7	20.7	0
104	0.0	738	75.3	18.7	0
115	0.0	1,190	75.8	23.0	0
117	0.0	650	76.8	17.6	0
118	0.0	548	77.0	15.0	0
123	0.0	640	76.8	16.9	0
125	0.0	790	74.1	25.7	0

Location/Room	CO (ppm)	CO <sub>2</sub> (ppm)	Temperature (°F)	Relative Humidity (%)	VOC (ppb)
<b>Lake View Campus</b>					
217	0.0	620	74.7	19.8	0
220	0.0	642	75.5	18.1	0
216	0.0	755	76.4	16.3	0
215	0.0	560	76.1	14.4	0
216b	0.0	607	75.5	16.6	0
215b	0.0	621	75.8	15.8	0
214	0.0	576	75.6	14.3	0
209	0.0	1,257	76.0	22.4	0
210	0.0	705	74.3	13.9	0
204	0.0	621	73.9	17.5	0
203	0.0	1,005	73.7	25.2	0
201	0.0	642	73.5	18.4	0
200	0.0	554	73.4	14.0	0
Exterior	0.0	445	66.4	10.8	0

**Table B-2**  
**Summary of Air Monitoring Date – Particle Counts**

Location/Room	Particle Size					
	0.3 um	0.5 um	1.0 um	2.0 um	5.0 um	10.0 um
Particle Count Range (Count/L)						
<b>Mountain View Campus</b>						
Old Library	9,514	2,012	1,012	723	235	52
Kitchen/Cafeteria	18,858	6,252	2,017	800	49	9
Crawlspace	12,865	4,527	2,773	2,183	900	235
Gym/Locker/Weight	11,206	1,692	408	191	44	13
112	11,408	1,961	681	430	150	48
120	10,702	1,943	599	306	80	24
217	23,538	4,743	1,278	754	189	48
218	29,783	5,814	1,212	713	177	35
210	28,652	4,370	604	263	48	12
Second Floor Hall	31,170	5,611	1,066	581	144	38
226	10,542	2,129	807	455	92	21
116	10,925	2,775	1,272	813	223	45
Principal	9,894	1,605	444	240	60	17
507	4,733	870	259	143	34	10
Front Entrance	10,950	1,598	294	81	10	3
Gym Entrance	10,565	1,403	254	65	6	2
<b>Lake View Campus</b>						
Library 121	13,665	3,516	1,668	1,077	296	69
Boiler	12,371	2,291	703	361	86	25
Shop 003	10,770	1,579	327	115	12	2
Music 017	12,694	3,271	1,519	959	287	88
025	12,267	2,258	623	252	31	8
Cafeteria	19,783	7,632	3,536	1,424	73	12
004	16,798	5,675	2,571	1,030	62	15
New Gym	11,016	1,413	257	92	15	4
001 OT/PT	11,061	1,472	277	113	19	4
007	9,749	1,885	715	433	115	32
009	9,658	1,723	619	369	115	30
100	12,371	1,883	552	287	77	20
102	12,445	3,518	1,899	1,291	475	145
101	12,208	2,998	1,265	594	93	24
103	14,568	2,537	890	355	30	7
Gym 105	11,760	2,419	909	412	49	11
113	11,372	2,434	976	473	85	24
104	10,228	1,659	551	298	87	35
115	8,898	2,091	960	590	152	34
117	10,480	1,970	727	348	66	20
118	9,860	1,567	478	222	41	8
123	10,424	1,993	733	355	51	14
125	11,700	2,015	651	336	84	22
217	11,614	2,268	857	391	47	9
220	10,317	2,163	914	467	86	25

Location/Room	Particle Size					
	0.3 um	0.5 um	1.0 um	2.0 um	5.0 um	10.0 um
Particle Count Range (Count/L)						
<b>Lake View Campus</b>						
216	10,033	2,197	926	463	73	20
215	9,840	1,779	608	264	25	7
216b	11,700	2,956	1,275	568	44	12
215b	12,661	3,679	1,671	706	34	10
214	10,471	2,615	1,173	521	51	18
209	10,622	3,642	2,012	1,107	223	47
210	11,274	2,9927	1,373	636	81	23
204	22,273	10,915	5,7890	2,373	64	15
203	19,412	6,314	3,047	1,409	150	37
201	15,119	5,775	2,921	1,239	52	13
200	17,424	7,305	3,879	1,602	32	7
Exterior	9,472	1,073	264	96	12	4

## **APPENDIX C**

### **SUMMARY OF LABORATORY ANALYSIS RESULTS**



**Table C-1  
 Summary of Mold Spore Count Analysis Results  
 Mountain View Campus  
 7530 Court Street – Elizabethtown, New York  
 Air Samples Collected on April 25, 2024**

Spore Count Concentration (count/m <sup>3</sup> )										
Sample Type	Interior Area of Concern								Exterior Background	
Sample ID	MA01	MA02	MA03	MA04	MA05	MA06	MA07	MA08	MA09	MA10
Sample Location	Old Library	Crawlspace	Gym	Room No. 112	Room No. 120	Room No. 217	Room No. 210	Principal Office	Front Entrance	Gym Entrance
Mycelial Fragments	40	<b>610</b>	<b>93</b>	<b>120</b>	53	<b>120</b>	<b>150</b>	67	67	27
Pollen	<13	<13	27	<13	13	13	13	<13	13	53
Total Fungal Spores	170	<b>1,000</b>	250	<b>530</b>	<b>560</b>	<b>750</b>	<b>510</b>	270	360	280
Alternaria	<13	<13	<13	<13	<13	<13	<13	<b>13</b>	<13	<13
Ascospores	<13	27	13	13	<13	13	27	<13	27	13
Aspergillus/ Penicillium-like	27	<b>230</b>	<b>53</b>	27	<b>93</b>	<b>160</b>	13	27	27	13
Basidiospores	67	120	110	170	<b>250</b>	<b>240</b>	170	93	190	200
Bipolaris/ Drechslera	<13	<13	<13	<13	<13	<13	<13	<13	<13	<13
Chaetomium	<13	<b>270</b>	<13	<13	<13	<13	<13	<13	<13	<13
Cladosporium	27	<b>93</b>	<b>53</b>	<b>160</b>	<b>67</b>	<b>120</b>	<b>250</b>	<b>80</b>	27	13
Curvularia	<13	<13	<13	<b>13</b>	<13	<b>13</b>	<13	<13	<13	<13
Rusts/Smuts	<13	<b>80</b>	<13	<b>80</b>	27	53	27	13	53	27
Stachybotrys	<13	<13	<13	<13	<13	<13	<13	<13	<13	<13
Other/Unidentified	<b>53</b>	<b>190</b>	27	<b>67</b>	<b>120</b>	<b>150</b>	13	40	40	13

*Notes:*  
 Bold concentration for any of the interior area of concern air samples indicates a detectable presence of a parameter greater than the highest of the corresponding concentrations reported for the background air samples.

**Table C-2  
 Summary of Mold Spore Count Analysis Results  
 Lake View Campus  
 25 Sisco Street – Westport, New York  
 Air Samples Collected on April 26, 2024**

Spore Count Concentration (count/m <sup>3</sup> )									
Sample Type	Interior Area of Concern								Exterior Background
Sample ID	MA11	MA12	MA13	MA14	MA15	MA16	MA17	MA18	MA19
Sample Location	Room No. 121	Cafeteria	Room No. 001	Room No. 101	Room No. 125	Room No. 217	Room No. 214	Room No. 203	Front Entrance
Mycelial Fragments	<b>110</b>	27	<b>93</b>	<b>93</b>	27	<b>40</b>	<b>170</b>	<b>190</b>	27
Pollen	<b>13</b>	<13	<b>13</b>	<b>13</b>	<b>13</b>	<13	<b>40</b>	<b>27</b>	<13
Total Fungal Spores	<b>360</b>	<b>370</b>	<b>310</b>	<b>470</b>	170	250	<b>390</b>	<b>610</b>	280
Alternaria	<13	<13	<13	<13	<13	<13	<13	<13	<13
Ascospores	13	<13	27	40	13	13	<13	13	40
Aspergillus/ Penicillium-like	13	<b>67</b>	13	<b>80</b>	<b>53</b>	40	13	<b>80</b>	40
Basidiospores	<b>240</b>	<b>210</b>	<b>240</b>	<b>190</b>	93	150	<b>210</b>	<b>230</b>	160
Bipolaris/ Drechslera	<13	<13	<13	<13	<13	<13	<13	<13	<13
Chaetomium	<13	<13	<13	<13	<13	<13	<13	<13	<13
Cladosporium	<b>40</b>	<b>27</b>	13	<b>110</b>	13	<b>40</b>	<b>130</b>	<b>80</b>	13
Curvularia	<13	<13	<13	<13	<13	<13	<13	<13	<13
Rusts/Smuts	<13	13	<13	13	<13	<13	13	<b>53</b>	13
Stachybotrys	<13	<13	<13	<13	<13	<13	<13	<13	<13
Other/Unidentified	<b>53</b>	<b>53</b>	13	<b>40</b>	<13	13	13	<b>160</b>	13

*Notes:*  
 Bold concentration for any of the interior area of concern air samples indicates a detectable presence of a parameter greater than the corresponding concentration reported for the background air sample.